

American Journal of Essential Oils and Natural Products

Available online at www.essencejournal.com

American Journal of Essential Oils and Natural Products

J

Е

0

N

р

ISSN: 2321 9114 AJEONP 2016; 4(1): 20-22 © 2016 AkiNik Publications Received: 16-11-2015 Accepted: 19-12-2015

Ali G Al-kaf

Pharmacognosy & Medicinal Chemistry Department, Faculty of Pharmacy, Sana'a University, P. O Box 18084, Sana'a Yemen

Rebecca A Crouch

Department of Chemistry, University of Alabama in Huntsville, Huntsville, AL 35899, USA

Annika Denkert

Leibniz Institute of Plant Biochemistry Halle, Department of Bioorganic Chemistry, Weinberg 306120 HALLE (SAALE) Germany

Andrea Porzel

Leibniz Institute of Plant Biochemistry Halle, Department of Bioorganic Chemistry, Weinberg 306120 HALLE (SAALE) Germany

Othman SS Al-Hawshabi

Biology Department, Faculty of Science, Aden University, Aden, P.O. Box 6235, Yemen

Nasser A Awadh Ali*

A) Pharmacognosy & Medicinal Chemistry Department, Faculty of Pharmacy, Sana'a University, P. O Box 18084, Sana'a Yemen
B) Pharmacognosy Department, Faculty of Clinical Pharmacognosy, Al-Baha University, Albaha, P.O. Box 1988, KSA

William N Setzer

Department of Chemistry, University of Alabama in Huntsville, Huntsville, AL 35899

Ludger Wessjohann

Leibniz Institute of Plant Biochemistry Halle, Department of Bioorganic Chemistry, Weinberg 306120 HALLE (SAALE) Germany

*Correspondence:

Nasser A Awadh Ali A) Pharmacognosy & Medicinal Chemistry Department, Faculty of Pharmacy, Sana'a University, P. O Box 18084, Sana'a Yemen B) Pharmacognosy Department, Faculty of Clinical Pharmacognosy, Al-Baha University, Albaha, P.O. Box 1988, KSA

Chemical composition and biological activity of essential oil of *Chenopodium ambrosioides* from Yemen

Ali G Al-kaf, Rebecca A Crouch, Annika Denkert, Andrea Porzel, Othman SS Al-Hawshabi, Nasser A Awadh Ali*, William N Setzer, Ludger Wessjohann

Abstract

The chemical composition of the hydrodistilled leaf essential oil from *Chenopodium ambrosioides* L. growing wild in Yemen was determined by GC-MS analysis, and its cytotoxic, and general antioxidant potential were evaluated. Major compounds of *C. ambrosioides* oil were ascaridole (54.2%), isoascaridole (27.7%) and *p*-cymene (8.1%). At concentrations of 50 and 25 µg/mL, the essential oil showed cytotoxic activity against HT29 (human colon adenocarcinoma cells), with growth inhibition of 100 and 56% (\pm 3). The free radical scavenging ability of the oil was assessed by the DPPH assay to show antiradical activity with IC₅₀ of 10.4 µg/mL. TLC-bioautographic assay was used to identify the acetylcholinesterase inhibitory effect, and ascaridole was isolated and characterized (ESIMS, ¹H NMR, ¹³C NMR and HMBC) as the responsible constituent for anticholinesterase activity.

Keywords: C.ambrosioides; GC-MS, ascaridole, antioxidant, cytotoxic.

1. Introduction

The genus *Chenopodium* (Chenopodiaceae) includes varieties of weedy herbs (more than 200 species) native to much of Europe, Asia including India and China, and both North and South America^[1] and eight of which are found in Yemen^[2].

C. ambrosioides L.is widely used in popular medicine as a vermifuge, emmenagogue and abortifacient ^[3]. *C. ambrosioides* acts as a diuretic and an anthelmintic, and it is used to treat wounds, respiratory problems, inflammatory and painful processes, bronchitis, tuberculosis, and rheumatism ^[4]. Externally, it has been used as a wash for hemorrhoids, as a poultice to detoxify snake bites and other poisons and is thought to have wound-healing properties ^[5]. The essential oil of *C. ambrosioides* (EOCA) is known to inhibit the growth of dermatophytes ^[6] and other filamentous fungi such as *Aspergillus, Fusarium*, and *Colletotrichum* ^[7]. EOCA possessed antiaflatoxigenic, antimalarial and antioxidant properties ^[4] as well as antihelmintica worm expelling activities ^[8].

There have been several reports on the essential oil composition of *C. ambrosioides*; EOCA has high levels of monoterpenes, mainly ascaridole, α -terpinene, *p*-cymene, and isoascaridole ^[7, 9-14]. At least seven different chemotypes have been identified: (1) ascaridole, (2) α -terpinene, (3) α -pinene, (4) *p*-cymene, (5) carvacrol, (6) α -terpinyl acetate, and (7) limonene ^[11]. To our knowledge, there are no published reports on the chemical composition, cytotoxic and antioxidant activity of *C. ambrosioides* oil from Yemen. For this reason, the chemical composition of EOCA was analyzed and its antiproliferative activity on a human colon adenocarcinoma cell line was investigated. The antioxidant potential of the oil was also evaluated by the chemical DPPH assay. A bioautographic assay was used to identify the acetylcholinesterase inhibitory activity. The active compound was isolated and characterized using ESIMS, LC-MS, ¹H NMR, ¹³C NMR and HMBC, and identified as ascaridole.

2. Materials and Methods

2.1 Plant Material

The leaves of *C. ambrosioides* were collected in the early morning from the Alselw district, Taiz province, Yemen, on 13th August 2012. The plant was identified by Dr. Hassan M. Ibrahim of the Botany Department, Faculty of Sciences, Sana'a University. A voucher specimen (YMP-chen-14) has been deposited at the Pharmacognosy Department, Sana'a University, Yemen.

2.2 Essential Oil Extraction

Dried leaves from *C. ambrosioides* were hydrodistilled for 3 h in a Clevenger type apparatus according to the European Pharmacopoeia ^[15]. The obtained oil was subsequently dried over anhydrous Na₂SO₄ and kept at 4 °C until analysis.

2.3 Gas Chromatography – Mass Spectral Analysis

The essential oil of *C. Ambrosioides* was analyzed by GC-MS according to method described in ^[16]. The essential oil composition of *C. ambrosioides* is summarized in Table 1.

2.4 Antioxidant Activity Screening

The antiradical activity of the oil was studied using DPPHassay as previously reported ^[16].

2.5 Cytotoxicity Screening

The cytotoxic activity of oil was studied against HT29 tumor cells (human colonic adenocarcinoma cells) as previously reported ^[16].

2.6 Modified TLC Bioautographic Assay for Acetylcholinesterase Inhibition

The test was carried out according to reference ^[17]. PTLC layers were developed with toluene: ethylacetate (93:7, v/v). The positive band was scratched out, eluted with methanol and purified by HPL Cusing an RP-C₁₈cartidge and subjected to ESI and LC-MS measurements.

3. Results and Discussions

EOCA was obtained by hydrodistillation with a yield of 0.52%, v/w, on a dry-weight basis. Eleven compounds, representing 94.1% of the oil, were identified. As can be seen in Table 1, the main constituents of the essential oil were oxygenated monoterpenoids and monoterpene hydrocarbons at concentrations of 85.3% and 8.8%, respectively. Ascaridole (54.2%), isoascaridole (27.7%) and *p*-cymene (8.1%) were the main compounds in the oil. Yemeni oil composition therefore is somewhat similar to the compositions described in previous reports on C. ambrosioides oil from Madagascar with ascaridole (41.8%), isoascaridole (18.1%), *p*-cymene (16.2%) ^[9], Brazil with ascaridole (61.4%), isoascaridole (18.6%) ^[7], China with ascaridole (29.7%), isoascaridole (13.0%), pcymene (12.7%) ^[18], cultivated C. ambrosioides from Uttarakhand, India with ascaridole (45.0%), isoascaridole (2.9%), *p*-cymene (27.1%), α-terpinene (8.3%) ^[19], and Cuba with ascaridole (30.5-47.1%), p-cymene (20.2-21.1%) αterpinene (17.0-20.7%)^[10]; but differed from those described in Nigeria by Owolabi et al. [11] [with α-terpinene (63.1%), pcymene (26.4%) and ascaridole (3.9%)], in wild C. ambrosioides from Himalayan India by Lohani et al. [19] [with α -terpinene (44.7%), *p*-cymene (21.3%) and ascaridole (17.9%)], and in southern India by Gupta et al. ^[12] [with α terpinene (63.6%) and p-cymene (19.5%)]. The composition of EOCA from Yemen places this plant in the ascaridole chemotype [11].

Table I: Essential oil composition of *Chenopodium ambrosioides* from Yemen.

RI	Compound	%
1017	α-Terpinene	0.7
1025	<i>p</i> -Cymene	8.1
1125	Unidentified	4.8
1185	<i>p</i> -Cymen-8-ol	tr
1224	Unidentified	0.8
1238	Ascaridole	54.2
1251	cis-Piperitone epoxide	0.4
1257	trans-Piperitone epoxide	1.2
1268	trans-Ascaridole glycol	tr
1288	cis-Ascaridole glycol	1.8
1295	Carvacrol	tr
1303	Isoascaridole	27.7
1316	4-Hydroxycryptone	tr
	Oxygenated monoterpenoids	85.3
	Monoterpene hydrocarbons	8.8
	Total Identified (11)	94.1

EOCA shows a potent radical scavenging activity at concentrations ranging from 20 to 5 μ g/mL. Its chemical antiradical activity with an IC₅₀ of 9.7 μ g/mL, thus is better than that of ascorbic acid (IC₅₀ 12.7 μ g/mL). EOCA is markedly rich in oxygenated monoterpenoids (85.3%) which may act as the radical-scavenging agents. It seems to be a general trend that the essential oils that contain oxygenated monoterpenoids have greater antioxidative properties than non-oxygenated ones ^[20].

The potential anticancer activity of EOCA was assessed against HT29 (human colon adenocarcinoma) cells. At concentrations of 50 and 25 μ g/mL, EOCA showed good cytotoxic activity, with growth inhibition of 100% and 56% (± 3.45), respectively. The potent antineoplastic properties of EOCA may be attributed to the main component of the oil, ascaridole which is known to exhibit activity against different tumor cell lines *in vitro* ^[21, 22].

EOs and their components with marked anticholinesterase

activity can find practical applications, for example, as natural insecticides, anti-dementia agents, and for treating head lice ^[23, 24]. The inhibitory effect of EOCA on AChE was detected by bioautographic TLC assay. The active band was isolated, purified and characterized by ESIMS, LC-MS, ¹H NMR, ¹³C NMR and HMBC as ascaridole.

4. Conclusions

In conclusion, EOCA is characterized by high content of oxygenated monoterpenes, mainly ascaridole and isoascaridole. The results clearly show that the oil possesses a significant cytotoxic activity against human tumor HT29 cell line. The cytotoxic activity can be explained partly by the presence of ascaridole. Our results show a good correlation between antioxidant and cytotoxic activity of EOCA. A further study is needed to determine the anticholinesterase activity of ascaridole.

5. References

- 1. Smith BD. Eastern North America as an independent center of plant domestication. Proceeding of the National Academy of Sciences of the United States of America 2006; 103:12223-12228.
- Wood JRI. A Handbook of the Yemen Flora. Whitstable Litho Printers Ltd, Royal Botanic Gardens, Kew. UK. 1997, 398.
- 3. Comway GA, Slocumb JC. Plants used as abortifacients and emmenagogues by Spanish New Mexicans. Journal of Ethnopharmacology. 1979; 1:241-261.
- Kumar R, Kumar MA, Dubey NK, Tripathi YB. Evaluation of *Chenopodium ambroisioides* oil as a potential source of antifungal, antiaflatoxigenic and antioxidant activity. International Journal of Food Microbiology. 2007; 115:159-164.
- Chevallier A. The Encyclopedia of Medicinal Plants. Dorling Kindersley, London, 1996.
- Kishore N, Chansouria JPN, Dubey NK. Antidermatophytic action of the essential oil of Chenopodium ambrosioides and an ointment prepared from it. India Phytotherapy Research 1999; 10:453-455.
- Jardim CM, Jham GN, Dhingra OD, Freire MM. Composition and antifungal activity of the essential oil of the Brazilian *Chenopodium ambrosioides* L. Journal of. Chemical Ecology. 2008; 34:1213-1218.
- 8. Potawale SE, Luniya KP, Mantri RA, Mehta UK, Waseem MD, Sadiq MD *et al. Chenopodium ambrosioides*: An ethnopharmacological review. Pharmacology Online 2008; 2:272-286.
- Cavalli JF, Tomi F, Bernardini AF, Casanova J. Combined analysis of the essential oil of *Chenopodium ambrosioides* by GC, GC-MS and ¹³C-NMR spectroscopy: quantitative determination of ascaridole, a heat-sensitive compound. Phytochemical Analysis 2004; 15:275-279.
- Monzote L, Nance MR, García M, Scull R, Setzer WN. Comparative chemical, cytotoxicity and antileishmanial properties of essential oils from *Chenopodium ambrosioides*. Natural Product Communications 2011; 6:281-286.
- 11. Owolabi MS, Lajide L, Oladimeji MO, Setzer WN, Palazzo MC, Olowu RA *et al.* Volatile constituents and antibacterial screening of the essential oil of *Chenopodium ambrosioides* L. growing in Nigeria. Natural Product Communications 2009; 4:989-992.
- Gupta D, Charles R, Mehta VK, Garg SN, Kumar S. Chemical examination of the essential oil of *Chenopodium ambrosioides* L. from the Southern Hills of India. Journal of Essential Oil Research. 2002; 14:93-94.
- 13. Kiuchi F, Itano Y, Uchiyama N, Honda G, Tsubouchi A, Nakajima-Shimada J *et al.* Monoterpene hydroperoxides with trypanocidal activity from *Chenopodium ambrosioides*. Journal of Natural Products. 2002; 65:509-512.
- MacDonald D, VanCrey K, Harrison P, Rangachari PK, Rosenfeld J, Warren C *et al.* Ascaridole-less infusions of *Chenopodium ambrosioides* contain a nematicide (s) that is(are) not toxic to mammalian smooth muscle. Journal of Ethnopharmacology. 2004; 92:215-221.
- 15. Council of Europe. European Pharmacopoeia, 3rd Ed., Council of Europe Press, Strasbourg, 1997, 121-122.
- 16. Ali AAN, Sharopov FS, Al-kaf AG, Hill GM, Arnold N, Al-Sokari SS *et al.* Composition of essential oil from

Tagetes minuta and its cytotoxic, antioxidant and antimicrobial activities. Natural Product Communications 2014; 9:265-268.

- 17. Yang Z, Zhang X, Duan D, Song Z, Yang M, Li S. Modified TLC bioautographic method for screening acetylcholinesterase inhibitors from plant extracts. Journal of Separation Sciences. 2009; 32:3257-3259.
- Chu SS, Hu JF, Liu ZL. Composition of essential oil of Chinese *Chenopodium ambrosioides* and insecticidal activity against maize weevil, Sitophilus zeamais. Pest Management Science 2011; 67:714-718.
- 19. Lohani H, Chauhan NK, Haider KKSZ, Andola HC. Comparative aroma profile of wild and cultivated *Chenopodium ambrosioides* L. from Uttarakhand. Journal of Essential Oil Bearing Plants. 2012; 15:657-661.
- Tepe B, Daferera D, Sokmen M, Polissiou M, Sokmen A. The in vitro antioxidant and antimicrobial activities of the essential oil and various extracts of *Origanum syriacum* L. var. bevanii. Journal of Science Food and Agriculture. 2004; 84:1389-1396.
- 21. Bezerra DP, Marinho Filho JD, Alves AP, Pessoa C, de Moraes MO, Pessoa OD *et al.* activity of the essential oil from the leaves of *Croton regelianus* and its component ascaridole. Chemistry Biodiversity 2009; 6:1224-1231.
- 22. Efferth T, Olbrich A, Sauerbrey A, Ross DD, Gebhart E, Neugebauer M. Activity of ascaridole from the anthelmintic herb *Chenopodium anthelminticum* L. against sensitive and multidrug – resistant tumor cells. Anticancer Research. 2002; 22C:4221-4224.
- Picollo MI, Toloza AC, MougabureCueto G, Zygadlo J, Zerba E. Anticholinesterase and pediculicidal activities of monoterpenoids. Fitoterapia 2008; 79:271-278.
- Houghton PJ, Ren Y, Howes MJ. Acetylcholinesterase inhibitors from plants and fungi. Natural Product Reports 2006; 23:181-199.